

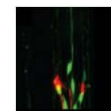
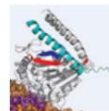
Tools for Mapping and Engineering Brain Computations

Ed Boyden

Synthetic Neurobiology Group
MIT

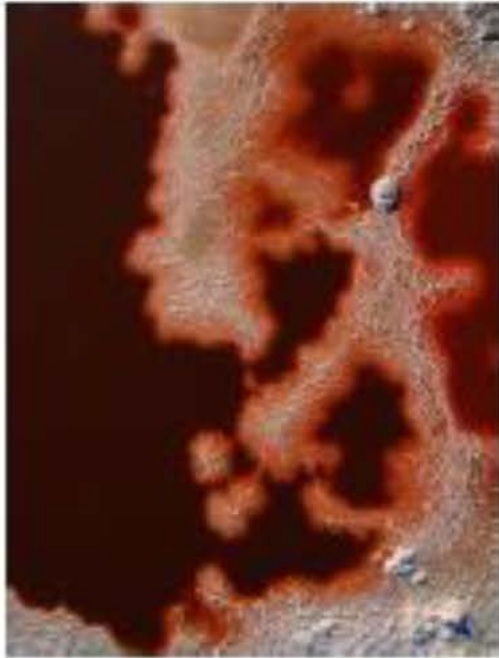


Biological Engineering



MIT CENTER FOR NEUROBIOLOGICAL ENGINEERING

Bacteriorhodopsins: Light-driven proton pumps



Courtesy of [Ricardo Mendonca Ferreira](#) on Flickr. CC license BY-NC-SA.

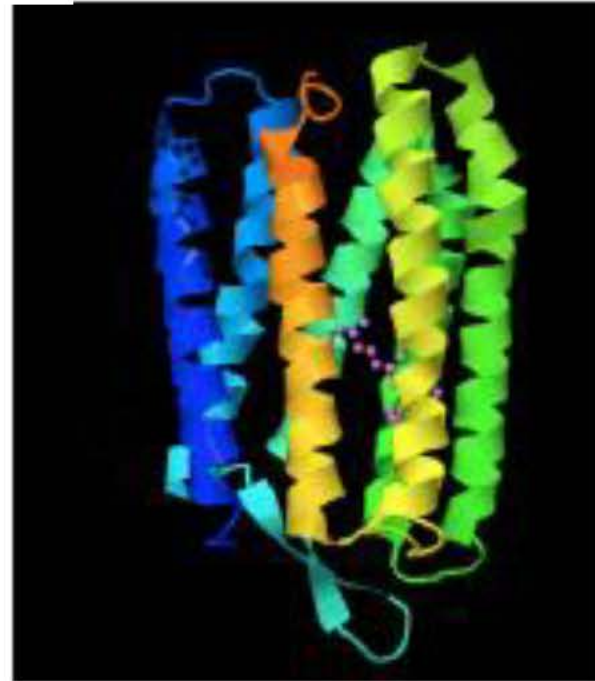


Image from the [RCSB PDB](#) of PDB ID 1DZE (Takeda, Kazuki, Yasuhiro Matsui, et al. "Crystal Structure of the M Intermediate of Bacteriorhodopsin: Allosteric Structural Changes Mediated by Sliding Movement of a Transmembrane Helix." *Journal of Molecular Biology* 341, no. 4 (2004): 1023–37.).

Oesterhelt, Dieter, and Walther Stoeckenius. "Rhodopsin-like Protein from the Purple Membrane of *Halobacterium Halobium*." *Nature* 233, no. 39 (1971): 149-52.



Halorhodopsins: Light-driven chloride pumps

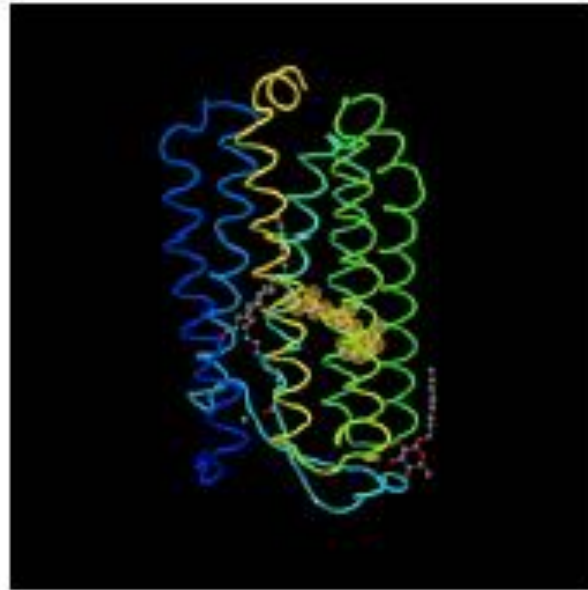


Image from the [RCSB PDB](#) of PDB ID 2JAF (Gmelin, Walter, Kornelius Zeth, et al. "The Crystal Structure of the L1 Intermediate of Halorhodopsin at 1.9 Å Resolution." *Photochemistry and Photobiology* 83, no. 2 (2007): 369–77.).

Matsuno-Yagi, Akemi, and Yasuo Mukohata. "Two Possible Roles of Bacteriorhodopsin; A Comparative Study of Strains of *Halobacterium Halobium* Differing in Pigmentation." *Biochemical and Biophysical Research Communications* 78, no. 1 (1977): 237-43.

Matsuno-Yagi, Akemi, and Yasuo Mukohata. "ATP Synthesis Linked to Light-dependent Proton Uptake in a Red Mutant Strain of *Halobacterium* Lacking Bacteriorhodopsin." *Archives of Biochemistry and Biophysics* 199, no. 1 (1980): 297-303.

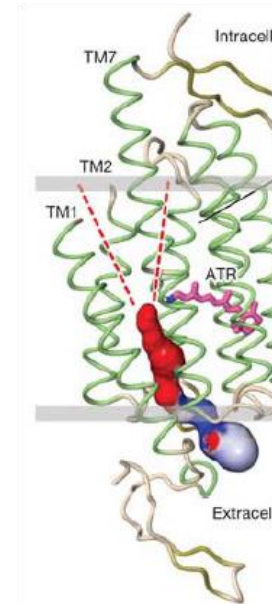
Schobert, Brigitte, and Janos K. Lanyi. "Halorhodopsin is a Light-driven Chloride Pump." *Journal of Biological Chemistry* 257, no. 17 (1982): 10306-13.



Channelrhodopsins: Light-driven cation pumps



Courtesy of [Proyecto Agua](#) on Flickr. CC license BY-NC-SA.



Reprinted by permission from Macmillan Publishers Ltd: Nature © 2012.

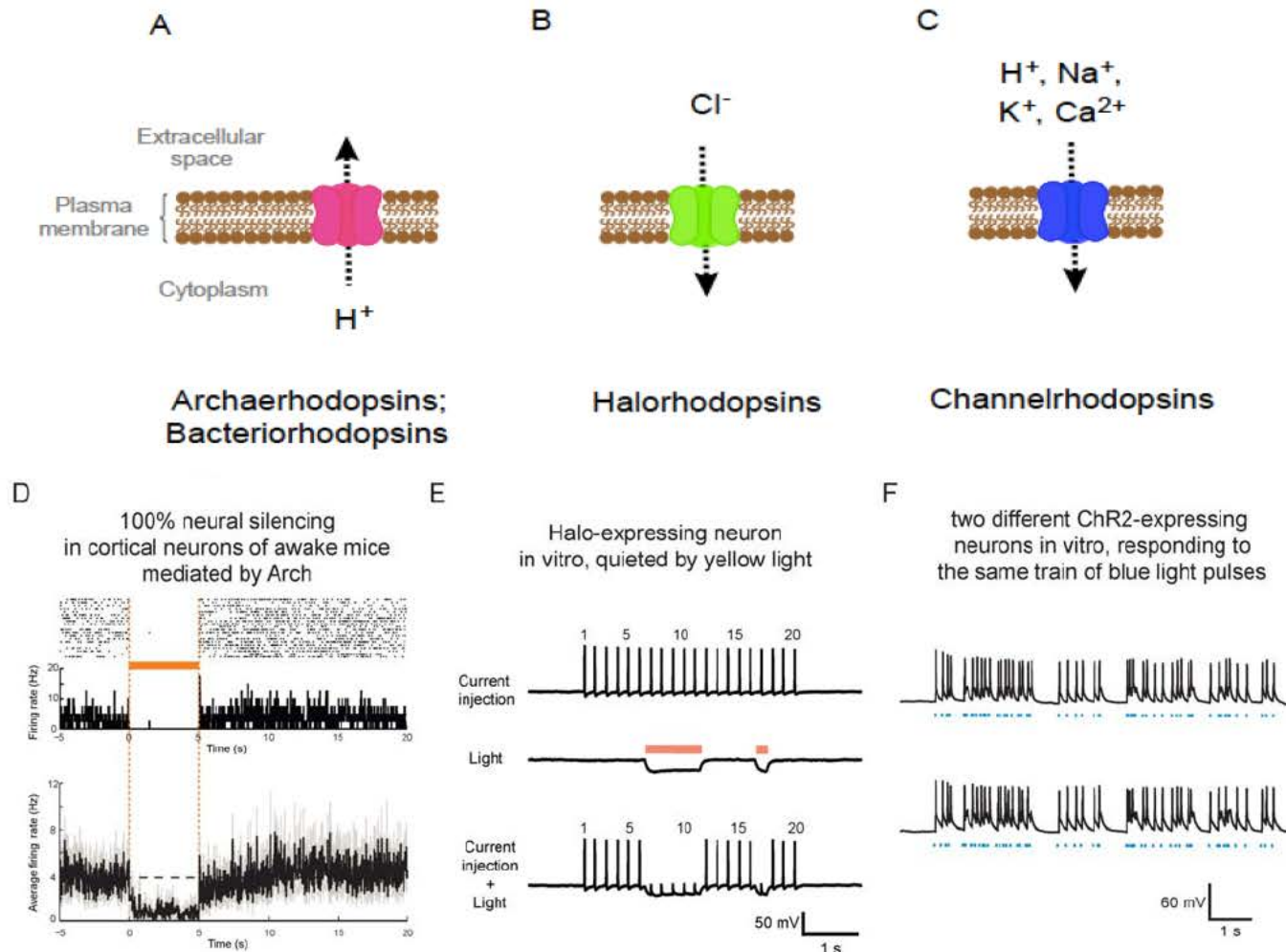
Source: Kato, Hideaki E., Feng Zhang, et al. "Crystal Structure of the Channel-rhodopsin Light-gated Cation Channel 482, no. 7385 (2012): 369–74.

Nagel, Georg, Doris Ollig, et al. "Channelrhodopsin-1: A Light-gated Proton Channel in Green Algae." *Science* 296, no. 5577 (2002): 2395-98.

Nagel, Georg, Tanjef Szellas et al. "Channelrhodopsin-2, a Directly Light-gated Cation-selective Membrane Channel." *Proceedings of the National Academy of Sciences* 100, no. 24 (2003): 13940-945.



Three major optogenetic molecule classes: microbial opsins, seven-transmembrane proteins,



Reprinted by permission from Macmillan Publishers Ltd: *Nature* © 2010.

Source: Chow, Brian Y., Xue Han, et al. "High-performance Genetically Targetable Optical Neural Silencing by Light-driven Proton Pumps." *Nature* 463, no. 7277 (2010): 98–102.

Courtesy of Han, Xue, and Edward S. Boyden. "Multiple-color Optical Activation, Silencing, and Desynchronization of Neural Activity, with Single-spike Temporal Resolution." *PloS One* 2, no. 3 (2007): e299. License CC BY.

Reprinted by permission from Macmillan Publishers Ltd: *Nature Neuroscience* © 2005. Source: Boyden, Edward S., Feng Zhang, et al. "Millisecond-timescale, Genetically Targeted Optical Control of Neural Activity." *Nature Neuroscience* 8, no. 9 (2005): 1263–68.



Proto-optogenetic experiments: heterologous expression of opsins in different cell types

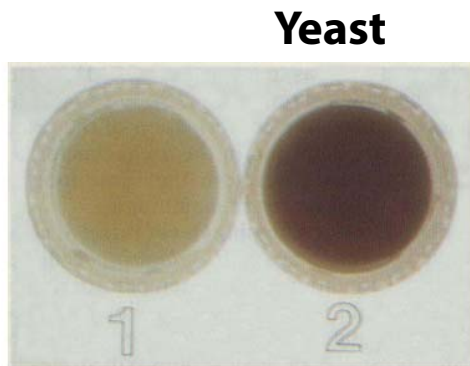


FIG. 1. Pelleted *S. pombe* cells. Pellet 1, cells transformed by the vector pEVP11, which lacks the *bop* gene. The yellow results from free retinal added to the culture medium. Pellet 2, cells transformed by the vector pEVBOp (pEVP11 containing the *bop* gene). The reddish color is a mixture of the purple of expressed bR and the yellow of free retinal.

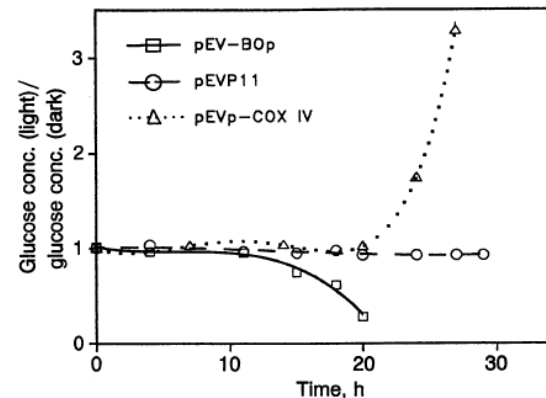


FIG. 5. Ratio of glucose concentrations in the growth medium for anaerobically growing *S. pombe* cultures with and without illumination [glucose conc. (light)/glucose conc. (dark)]. This ratio is plotted for three clones (pEV-BOp, pEVP11, and pEVp-COX IV). The absolute values of glucose concentrations are listed in Table 2.

Source: Hildebrandt, V., K. Fendler, et al. "Bacteriorhodopsin Expressed in Schizosaccharomyces Pombe Pumps Protons through the Plasma Membrane." *Proceedings of the National Academy of Sciences* 90, no. 8 (1993): 3578-82. Copyright © 1993 National Academy of Sciences, U. S. A.

Source: Hoffmann, Astrid, Volker Hildebrandt, et al. "Photoactive Mitochondria: In Vivo Transfer of a Light-driven Proton Pump into the Inner Mitochondrial Membrane of Schizosaccharomyces Pombe." *Proceedings of the National Academy of Sciences* 91, no. 20 (1994): 9367-71. Copyright © 1994 National Academy of Sciences, U. S. A.

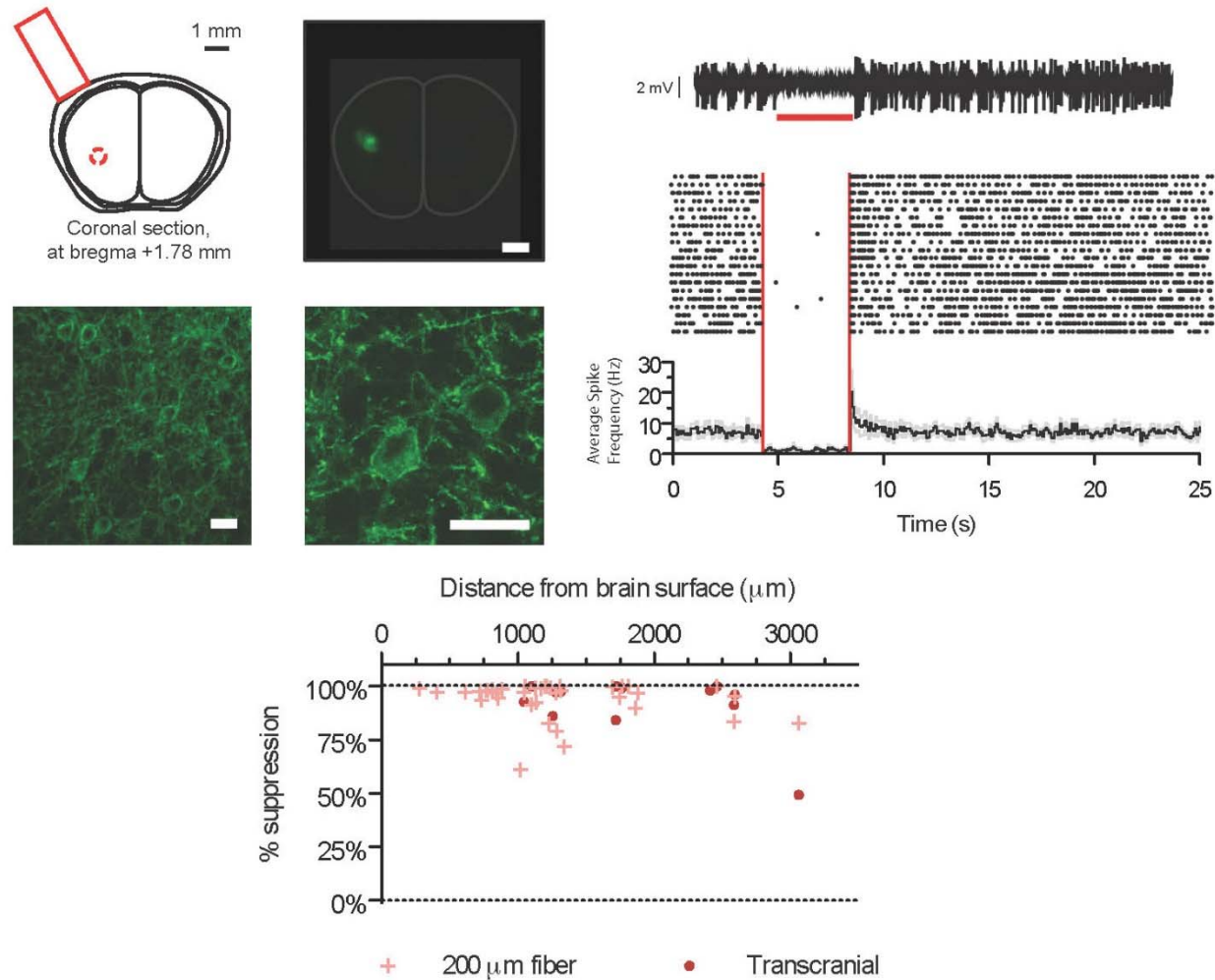
Hildebrandt V, Ramezani-Rad M, Swida U, Wrede P, Grzesiek S, Primke M, Büldt G. (1989) Genetic transfer of the pigment bacteriorhodopsin into the eukaryote *Schizosaccharomyces pombe*. *FEBS Lett.* 243(2):137-40.

Hildebrandt V, Fendler K, Heberle J, Hoffmann A, Bamberg E, Buldt G (1993) Bacteriorhodopsin expressed in *Schizosaccharomyces pombe* pumps protons through the plasma membrane. *Proc Natl Acad Sci U S A*, 90:3578-82.

Hoffmann A, Hildebrandt V, Heberle J, Büldt G (1994) Photoactive mitochondria: in vivo transfer of a light-driven proton pump into the inner mitochondrial membrane of *Schizosaccharomyces pombe*. *Proc Natl Acad Sci U S A*. 91(20):93.

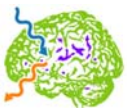


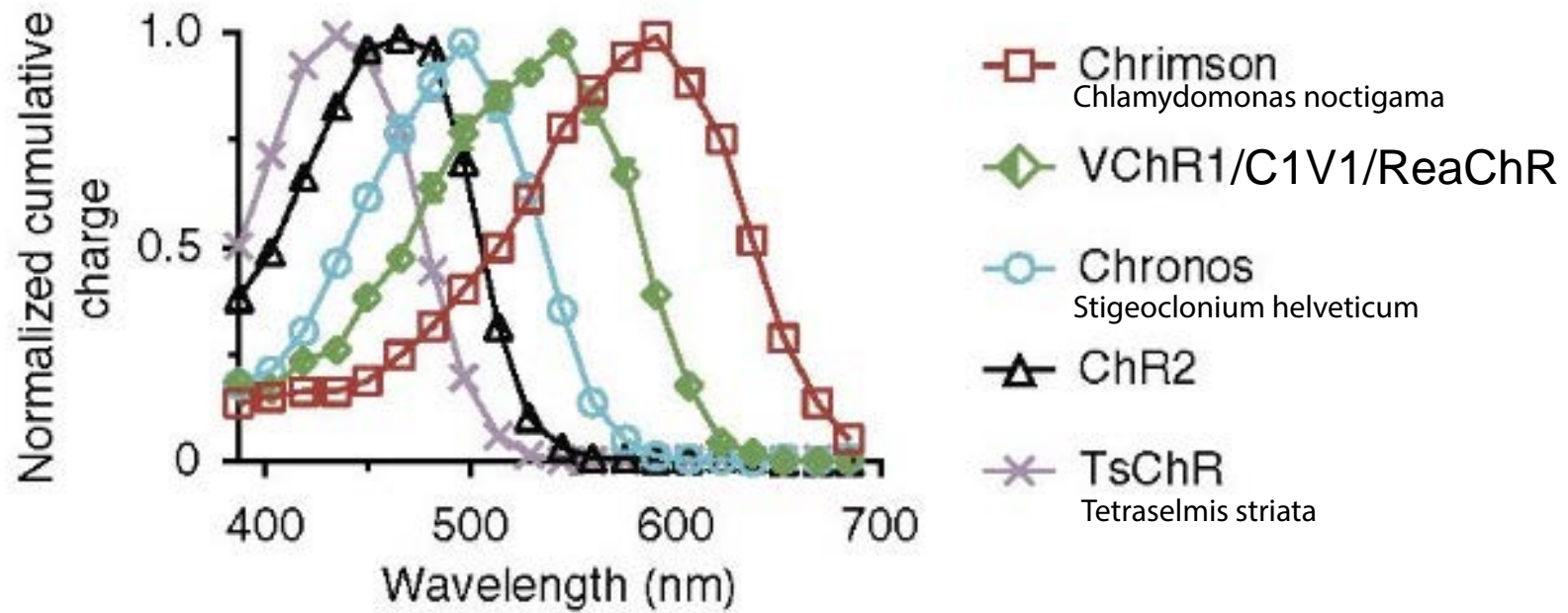
Noninvasive optogenetic neural silencing: Jaws



Chuong et al. (2014) *Nature Neuroscience*, doi:10.1038/nn.3752.

Reprinted by permission from Macmillan Publishers Ltd: *Nature Neuroscience* © 2014.
 Source: Chuong, Amy S., Mitra L. Miri, et al. "Noninvasive Optical Inhibition with a Red-shifted Microbial Rhodopsin." *Nature Neuroscience* 17, no. 8 (2014): 1123–29.



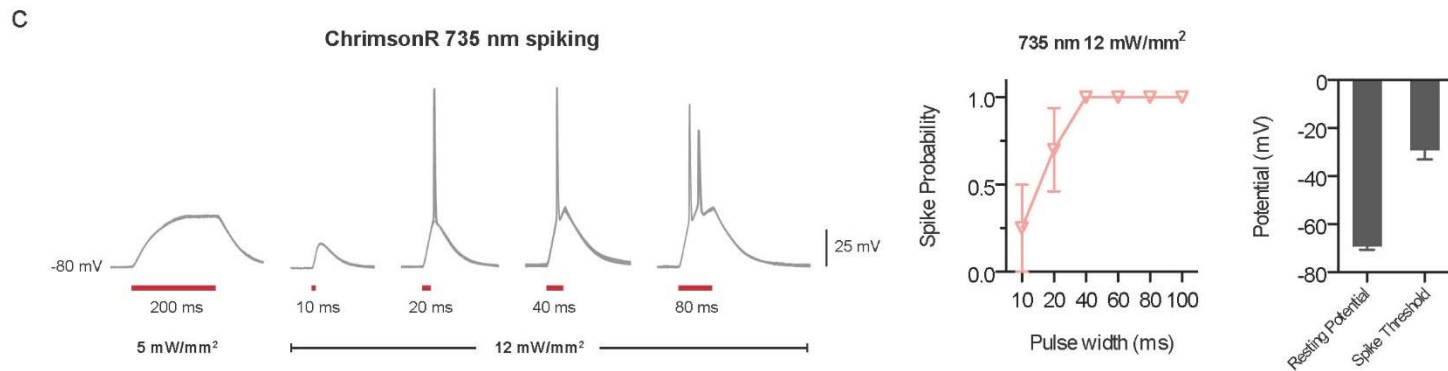
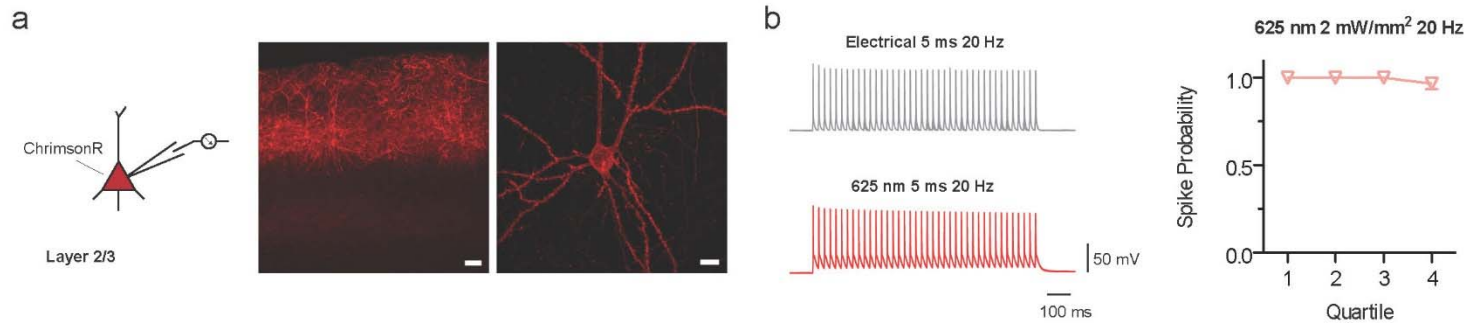
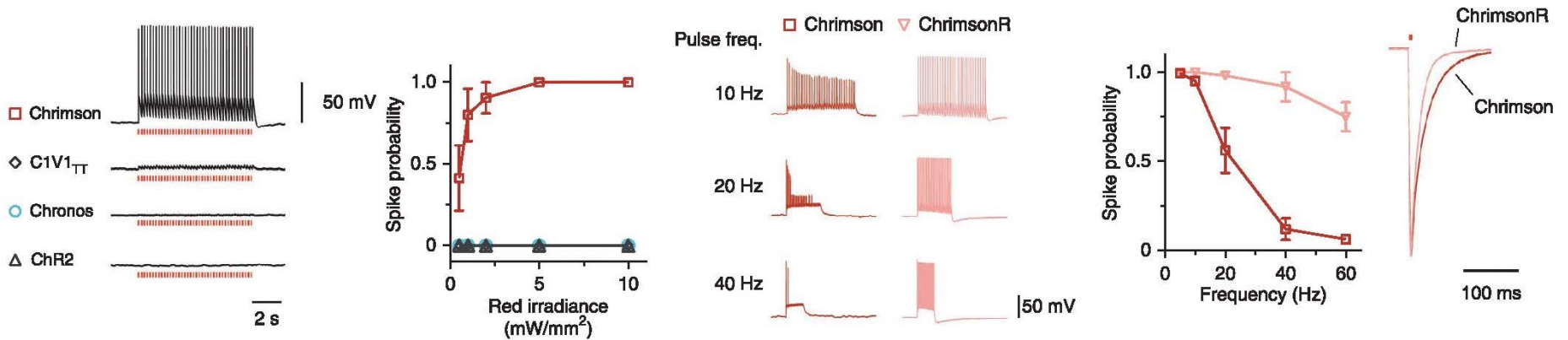


Reprinted by permission from Macmillan Publishers Ltd: *Nature Methods* © 2014.
 Source: Klapoetke, Nathan C., Yasunobu Murata, et al. "Independent Optical Excitation of Distinct Neural Populations." *Nature Methods* 11, no. 3 (2014): 338–46.

Klapoetke et al. (2014) *Nature Methods* 11:338–346.



Chrimson

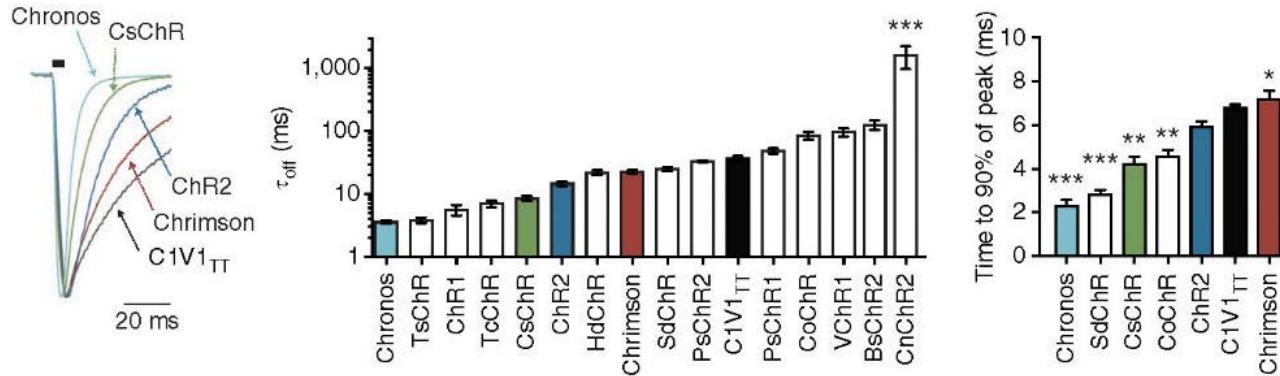


Klapoetke et al. (2014) *Nature Methods* 11:338–346.

Reprinted by permission from Macmillan Publishers Ltd: *Nature Methods* © 2014.
 Source: Klapoetke, Nathan C., Yasunobu Murata, et al. "Independent Optical Excitation of Distinct Neural Populations." *Nature Methods* 11, no. 3 (2014): 338–46.



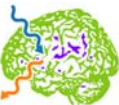
Chronos: a very fast channelrhodopsin...



Reprinted by permission from Macmillan Publishers Ltd: *Nature Methods* © 2014.
 Source: Klapoetke, Nathan C., Yasunobu Murata, et al. "Independent Optical Excitation of Distinct Neural Populations." *Nature Methods* 11, no. 3 (2014): 338–46.

Figures removed due to copyright restrictions.
 Please see supplemental figure 3 and figure 2A, B from Klapoetke, Nathan C., Yasunobu Murata, et al. "Independent Optical Excitation of Distinct Neural Populations." *Nature Methods* 11, no. 3 (2014): 338–46.
<http://www.ncbi.nlm.nih.gov/pmc/articles/PMC3943671/>

Klapoetke et al. (2014) *Nature Methods* 11:338–346.



...that is also very light sensitive!

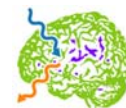
Figure removed due to copyright restrictions.

Please see figure 4C, D, E, F from Klapoetke, Nathan C., Yasunobu Murata, et al.

"Independent Optical Excitation of Distinct Neural Populations." *Nature Methods* 11, no. 3 (2014): 338–46.

<http://www.ncbi.nlm.nih.gov/pmc/articles/PMC3943671/>

Klapoetke et al. (2014) *Nature Methods* 11:338–346.

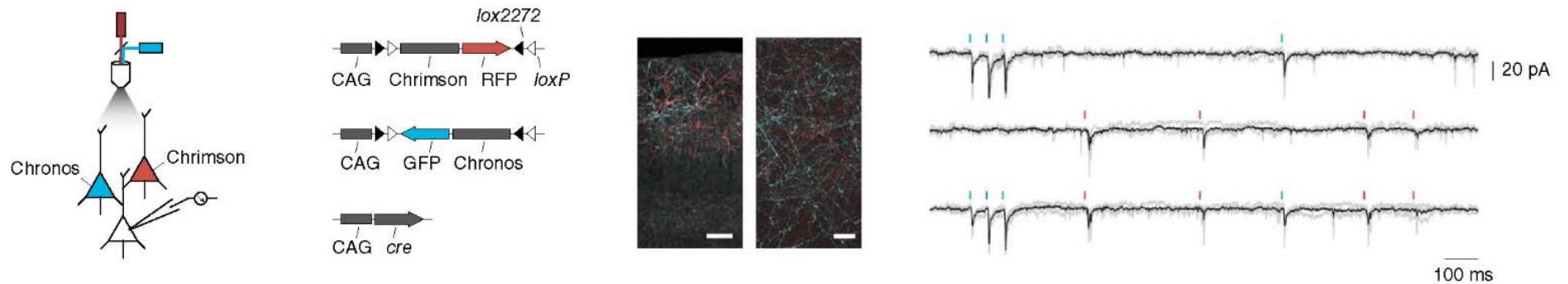


Chronos and Chrimson together

Figure removed due to copyright restrictions.

Please see supplemental figure 18 from Klapoetke, Nathan C., Yasunobu Murata, et al. "Independent Optical Excitation of Distinct Neural Populations." *Nature Methods* 11, no. 3 (2014): 338–46.

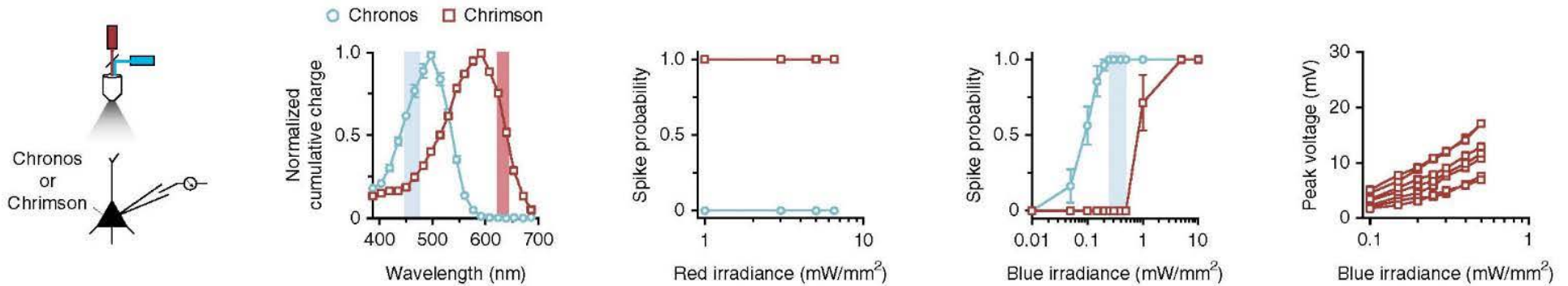
<http://www.ncbi.nlm.nih.gov/pmc/articles/PMC3943671/>



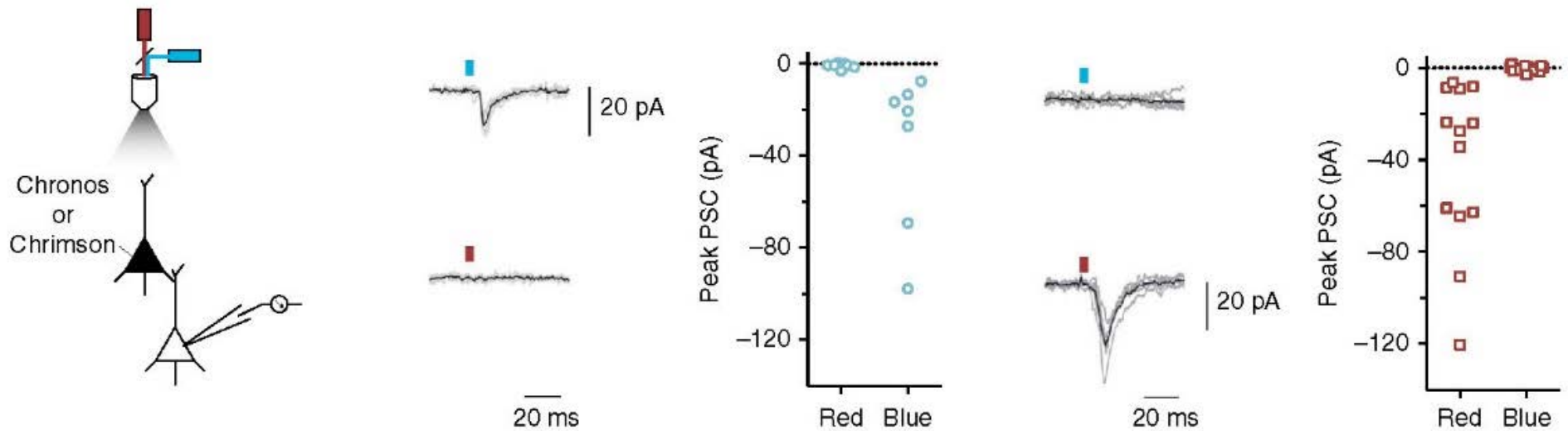
Klapoetke et al. (2014) *Nature Methods* 11:338–346.



Chronos and Chrimson together: zero-crosstalk control of spikes...



...and synaptic release events

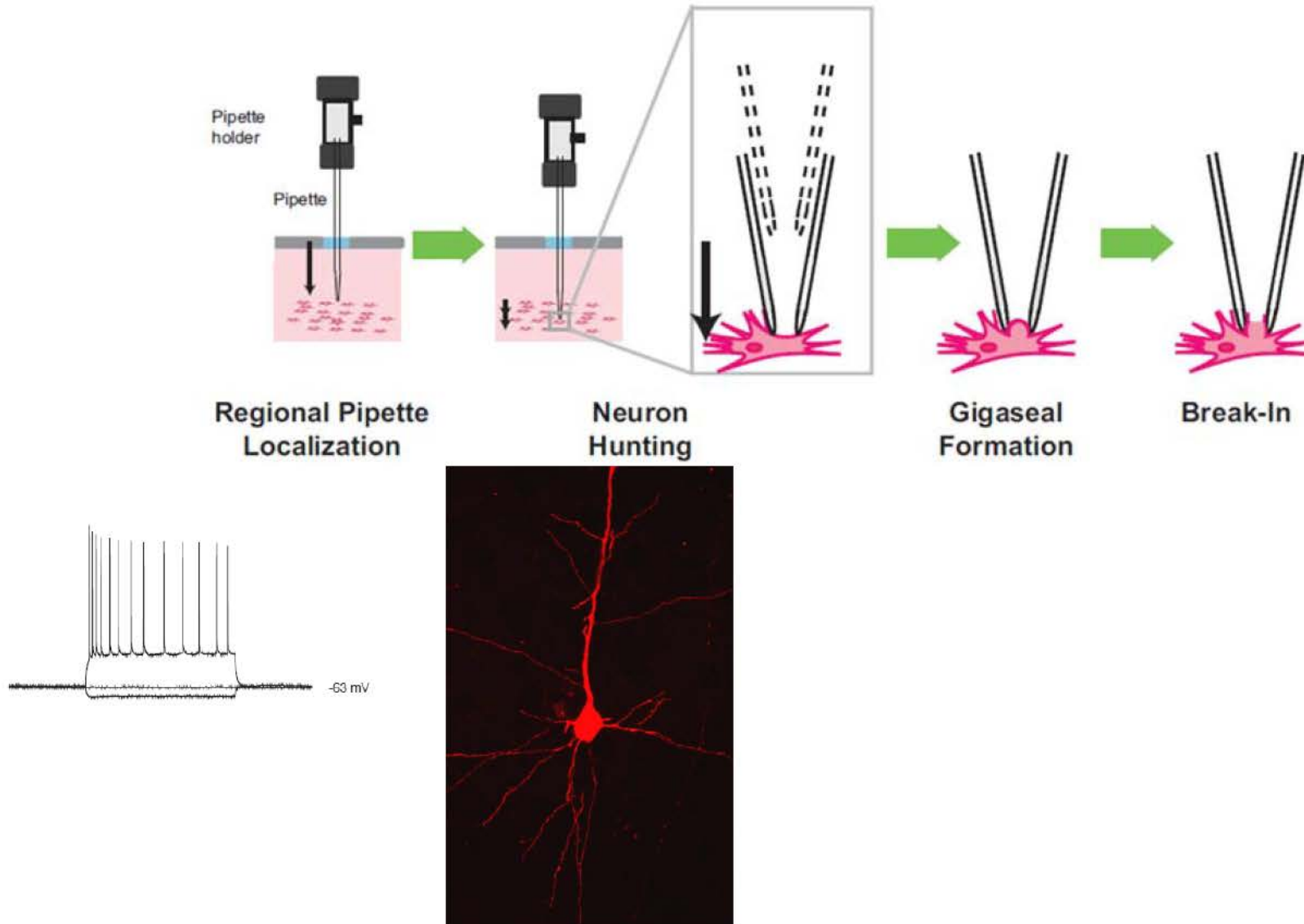


Klapoetke et al. (2014) *Nature Methods* 11:338–346.



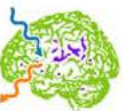
Reprinted by permission from Macmillan Publishers Ltd: *Nature Methods* © 2014.
 Source: Klapoetke, Nathan C., Yasunobu Murata, et al. "Independent Optical Excitation of Distinct Neural Populations." *Nature Methods* 11, no. 3 (2014): 338–46.

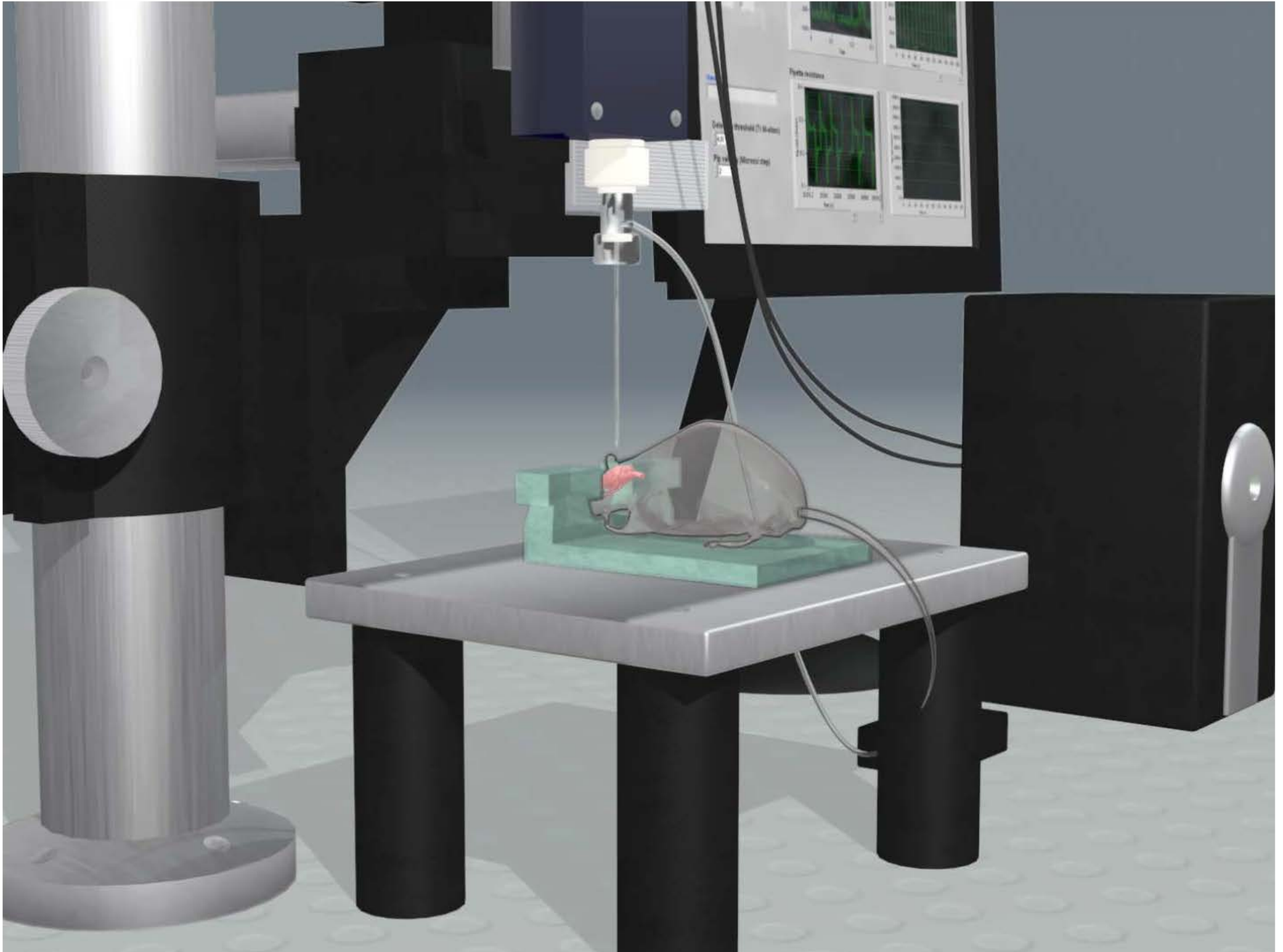
Whole cell patch clamp: enables simultaneous measurement of electrophysiology, morphology, and gene expression in single cells in living brain



Kodandaramaiah et al. (2012) *Nature Methods* 9:585–587.

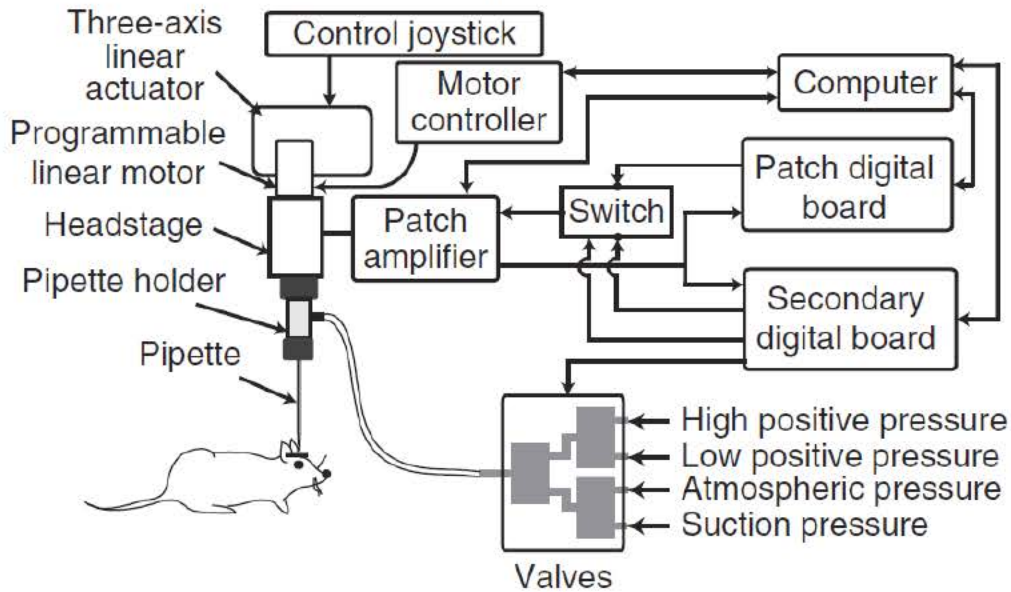
Reprinted by permission from Macmillan Publishers Ltd: *Nature Methods* © 2012.
Source: Kodandaramaiah, Suhasa B., Giovanni Talei Franzesi, et al. "Automated Whole-cell Patch-clamp Electrophysiology of Neurons in Vivo." *Nature Methods* 9, no. 6 (2012): 585–87.





Courtesy of The [McGovern Institute for Brain Research at MIT](#).
Used with permission. CC license BY-NC-SA.

A robot that can automatically patch clamp neurons in living brain

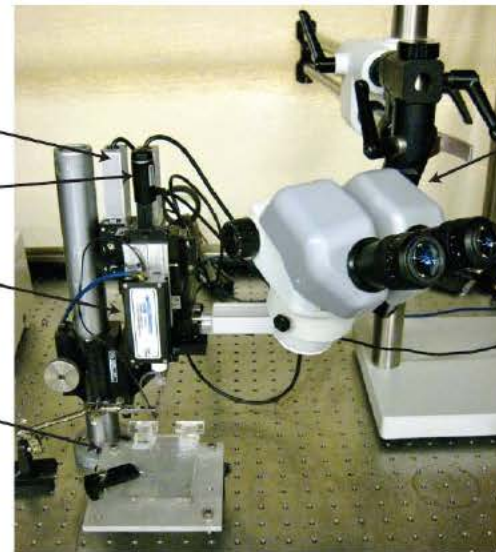


3 axis linear actuator
 MC1000e, Siskiyou

Programmable linear motor
 PZC12, Newport Inc

Headstage
 Molecular Devices

Low profile holder for head fixing the mouse



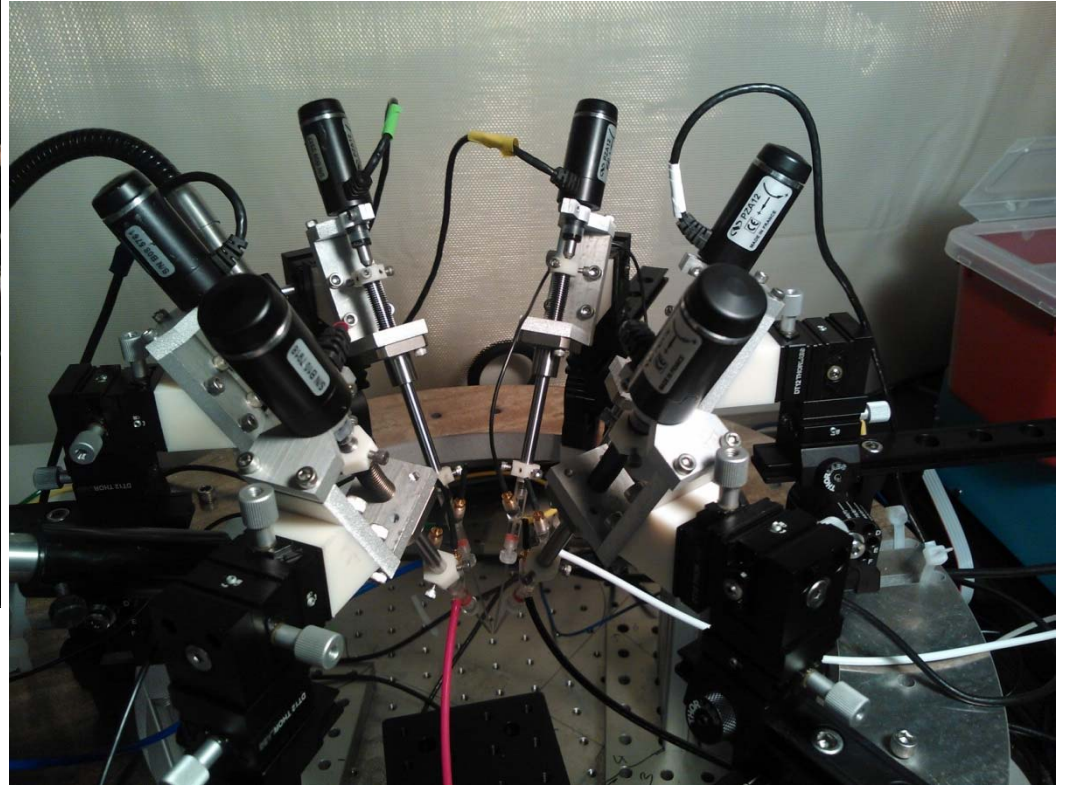
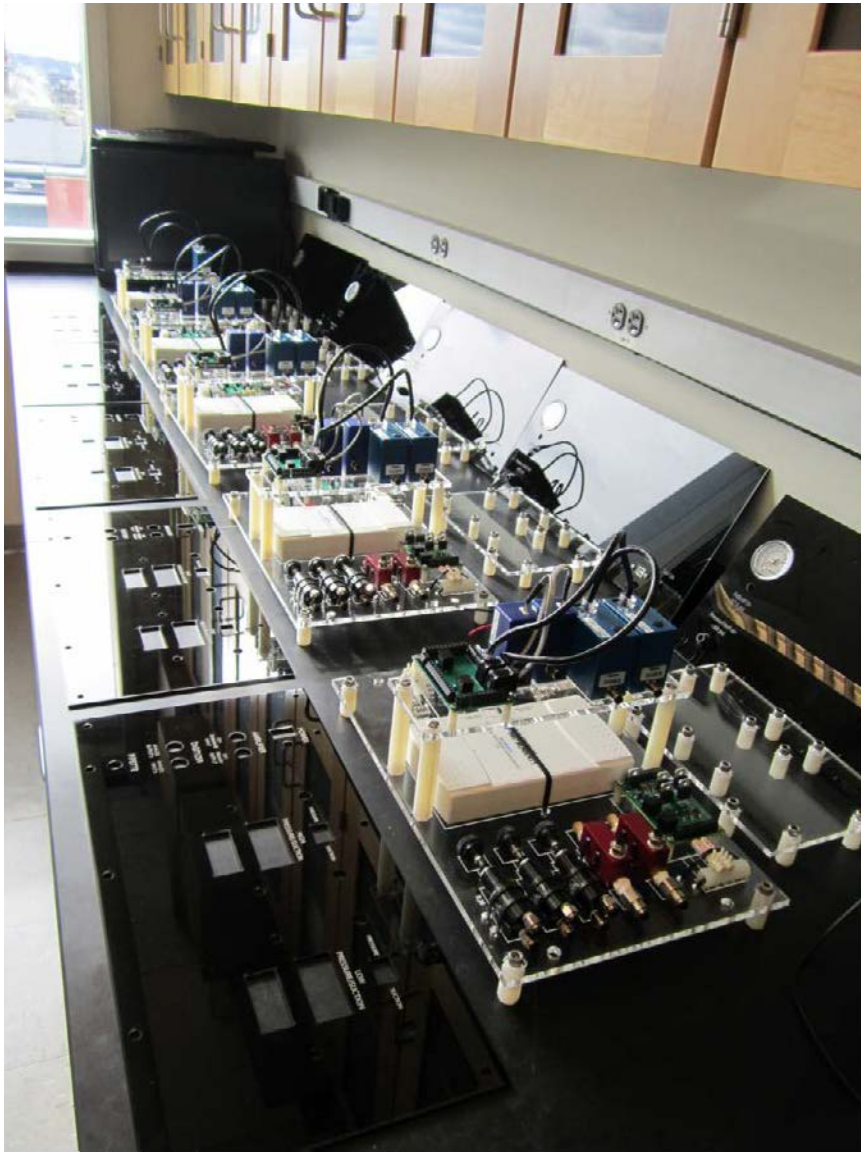
Stereo-microscope
 SM2616, Nikon

Reprinted by permission from Macmillan Publishers Ltd: *Nature Methods* © 2012.
 Source: Kodandaramaiah, Suhasa B., Giovanni Talei Franzesi, et al. "Automated Whole-cell Patch-clamp Electrophysiology of Neurons in Vivo." *Nature Methods* 9, no. 6 (2012): 585–87.

Kodandaramaiah et al. (2012) *Nature Methods* 9:585–587.

Commercialized by Neuromatic Devices, Inc. (ESB has no financial affiliation)

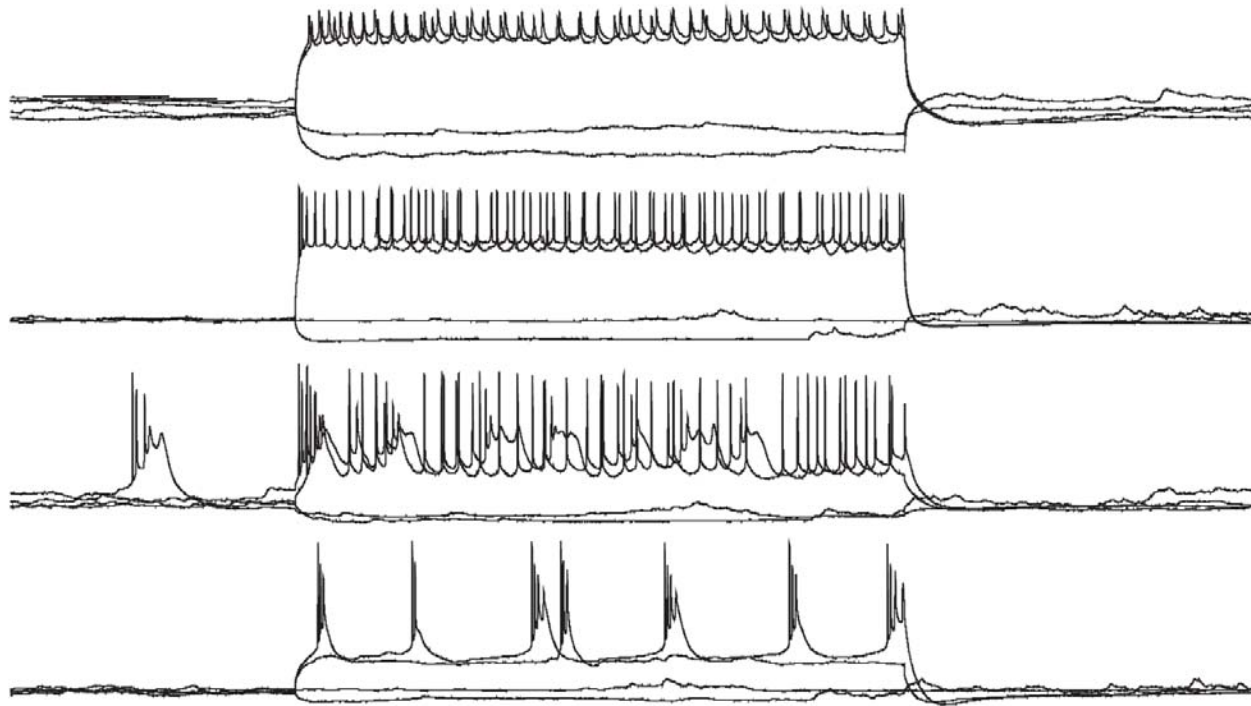




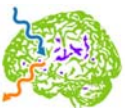
Suhasa Kodandaramaiah, Xue Han, Craig Forest



Robotic quad patching in living mouse brain



Suhasa Kodandaramaiah, Francisco Flores, Emery Brown, Craig Forest



Can we automate the rest of in vivo neuroscience?

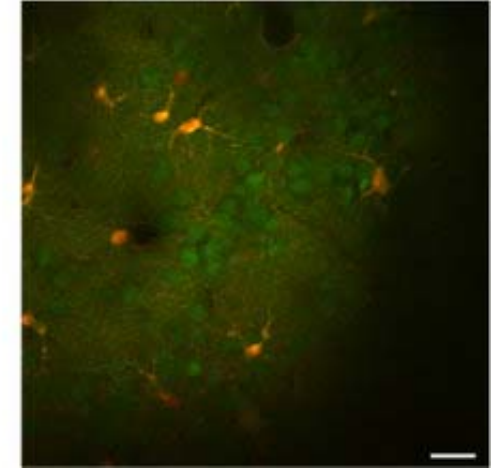
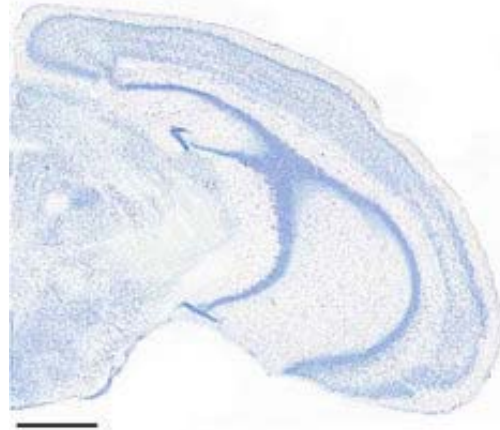
Figures removed due to copyright restrictions.
Please see Figures 1A, B and Figure 3B, C from Pak, Nikita, Joshua H. Siegle, et al. "Closed-loop, Ultraprecise, Automated Craniotomies." *Journal of Neurophysiology* (2015).



Figures removed due to copyright restrictions.
Please see Figures 1A, B and Figure 3B, C from Pak, Nikita, Joshua H. Siegle, et al. "Closed-loop, Ultraprecise, Automated Craniotomies." *Journal of Neurophysiology* (2015).

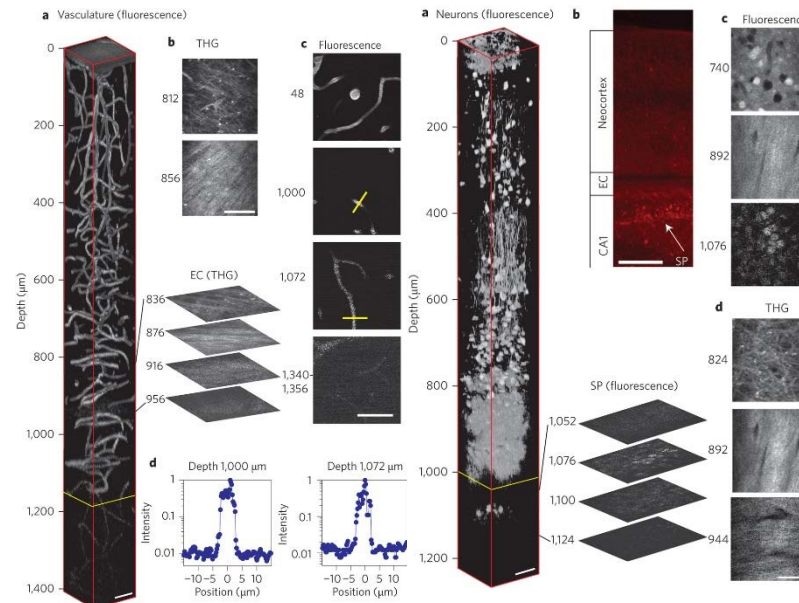


The world's smallest mammal: towards whole-organism functional imaging



Courtesy of [Bernard DUPONT](#) on Flickr. CC license BY-NC-SA.

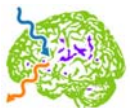
Claudia Roth-Alpermann and Michael Brecht (2009), [Scholarpedia](#), 4(11): 6830. CC license BY-NC-SA.



Reprinted by permission from Macmillan Publishers Ltd: *Nature Photonics* © 2013.

Source: Horton, Nicholas G., Ke Wang, et al. "In Vivo Three-photon Microscopy of Subcortical Structures within an Intact Mouse Brain." *Nature Photonics* 7, no. 3 (2013): 205–9.

**Michael Brecht, Ian Wickersham,
Susan Erdman**



Graduate Students, Postdocs, Staff

Adam Marblestone
Aimei Yang
Amy Chuong
Annabelle Singer
Anthony Zorzos
Asmamaw Wassie
Brian Allen
Caroline Moore-Kochlacs
Christian Wentz
Changyang Linghu
Daniel Martin-Alarcon
Daniel Schmidt
Demian Park
Desiree Dudley
Erica Jung
Fei Chen
Fumi Yoshida
Giovanni Talei Franzesi
Guangyu Xu
Harbaljit Sohal
Ho-Jun Suk
Ingrid van Welie
Ishan Gupta
Jae-Byum Chang
Jake Bernstein
Jorg Scholvin
Jun Deguchi
Justin Kinney
Kate Adamala
Kiryil Piatkevich
Kris Payer
Leah Acker
Lisa Lieberson
Manos Karagiannis
Mike Henninger
Nathan Klapoetke
Nikita Pak
Nir Grossman
Or Shemesh
Paul Tillberg
Ru Wang
Suhasa Kodandaramaiah
Yongxin Zhao
Young Gyu Yoon

Alumni

Alexander Guerra, Alex Rodriguez, Allison Dobry, Ash Turza, August Dietrich, Barbara Barry, Brian Chow (U Penn), Claire Ahn, Nate Greenslit (Harvard), Ian Wickersham (MIT), Ilya Kolb, Jenna Sternberg, Kyungman Kim, Masaaki Ogawa (NIPS), Masahiro Yamaguchi, Mike Baratta, Mingjie Li, Moshe Ben-Ezra, Rachel Bandler, Scott Arfin, Stephanie Chan, Sunanda Sharma, Tania Morimoto, Tim Buschman (Princeton), Victoria Wang, Xiaofeng Qian, Xue Han (BU), Yongku Cho (U. Conn.)

Undergraduate Students

Alexander Clifton, Bara Badwan, Deniz Aksel, Ellena Popova, Eunice Wu, Justine Cheng, Melina Tsitsiklis, Nico Enriquez, Rebecca Luoh, Semon Rezhnikov

Synthetic Neurobiology Group

<http://syntheticneurobiology.org/>

Funding

Allen Institute for Brain Science; AT&T; Bahaa Hariri; Benesse Corporation; Jerry and Marge Burnett; DARPA Living Foundries Program HR0011-12-C-0068; DARPA HR0011-11-14-0004; Department of Defense CDMRP PTSD Program; Google; Harvard/MIT Joint Grants Program in Basic Neuroscience; Human Frontiers Science Program; IET A. F. Harvey Prize; Joyce and Jeremy Wertheimer; Lincoln Labs Campus Collaboration Award; MIT Alumni Class Funds; MIT Intelligence Initiative; MIT McGovern Institute and McGovern Institute Neurotechnology (MINT) Program; MIT Media Lab and Media Lab Consortia; MIT Mind-Machine Project; MIT Neurotechnology Fund (& its generous donors); NARSAD; New York Stem Cell Foundation-Robertson Investigator Award; NIH Director's Pioneer Award 1DP1NS087724 and New Innovator Award 1DP2OD002002, NIH EUREKA Awards 1R01NS087950 and 1R01NS075421, NIH Transformative Awards 1R01MH103910 and 1R01GM104948, NIH Single Cell Grant 1R01EY023173, and NIH Grants 1R01DA029639, 1R43NS070453, 1RC2DE020919, 1RC1MH088182, 2R44NS070453, and 1R01NS067199; NSF INSPIRE Award CBET 1344219, NSF CAREER Award CBET 1053233 and NSF Grants, EFRI0835878, DMS0848804, DMS1042134 (the Cognitive Rhythms Collaborative), and CCF 1231216 (the Center for Brains Minds and Machines); Office of the Assistant Secretary of Defense for Research and Engineering; Paul Allen Distinguished Investigator in Neuroscience Award; Simons Foundation; Skolkovo Institute of Science and Technology; Alfred P. Sloan Foundation; Society for Neuroscience Research Award for Innovation in Neuroscience (RAIN); Stacy and Joel Hock; Synthetic Intelligence Project (& its generous donors); Wallace H. Coulter Foundation. Core grants: NIH P30-ES002109, 5 P30 EY002621-37 Pre-MIT: Hertz Foundation, Helen Hay Whitney Foundation.

Collaborating Groups

3-D Brain-building: Utkan Demirci
Blindness: Alan Horsager, Alapakkam Sampath, Bill Hauswirth, Botond Roska
C. elegans: Alipasha Vaziri, Manuel Zimmer
In vivo Robotics: Craig Forest, Hongkui Zeng
Microscopy: Alipasha Vaziri, Peter So, Ramesh Raskar
Neural modeling: Christoph Borgers, Fiona LeBeau, Miles Whittington, Nancy Kopell
Neural recording: George Church, Keith Tyo, Konrad Kording, Leaf labs
Opsin engineering: Adam Cohen, Beijing Genomics Institute, Ernst Bamberg, Gane Wong, Jess Cardin, Kay Tye, Martha Constantine-Paton, Michael Melkonian, Patrick Stern, Robert CampbellVivek Jayaraman, Yingxi Lin
Opto-fMRI: Ann Graybiel, Chris Moore, Itamar Kahn, Nancy Kopell, Randy Buckner
Optogenetic hardware: Clif Fonstad, Ferro Solutions Inc., Joseph Jacobson, Kendall Research Systems, Rahul Sarpeshkar, Steve Wasserman
Polymerase engineering: George Church, Keith Tyo, Konrad Koering
Primate work: Ann Graybiel, Bob Desimone, Bob Wurtz, Roderick Bronson, Wim Vanduffel
Transgenics: Hongkui Zeng



MIT OpenCourseWare
<http://ocw.mit.edu>

9.123 / 20.203 Neurotechnology in Action
Fall 2014

For information about citing these materials or our Terms of Use, visit: <http://ocw.mit.edu/terms>.