

Lecture notes courtesy of Wyan-Ching Mimi Lee. Used with permission.

Review Session

3/14/04

- $\text{Na}^+ \text{K}^+$ pump - can run many APs after turn off pump
- Nernst equilibrium: concentration gradient, electrical gradient give you battery

somatotopic - eg retinotopic cat on retina \rightarrow cat on visual cortex

local neighborhood relationships preserved (adjacent cells on retina project to adjacent cells in visual cortex)

eg tickle cells close together on body \rightarrow cells close together stimulated in brain

Nernst equation: $V = \frac{RT}{zF} \times \ln \frac{[I]_o}{[I]_i}$
 $= 58 \log \frac{[I]_o}{[I]_i}$ (at room temperature)

- good for any ions (takes care of its own sign)
- (in Goldman equation, negative ions have $[I]_i / [I]_o$, so can use same z for all terms)
- Goldman equation not covered enough to use on test

weighted-average equation: used for all H+H models

derived from Ohm's Law for Membranes

$$I = g_{\text{Na}} (V_m - E_{\text{Na}}) + g_{\text{K}} (V_m - E_{\text{K}})$$

$$0 = g_{\text{Na}} (V_m - E_{\text{Na}}) + g_{\text{K}} (V_m - E_{\text{K}})$$

$$V_m = \frac{g_{\text{Na}} E_{\text{Na}} + g_{\text{K}} E_{\text{K}}}{g_{\text{Na}} + g_{\text{K}}} \quad (\text{weighted by relative conductances})$$

- if only conductive to K^+ , $V_m \sim E_{\text{K}}$

- if only conductive to Na^+ , $V_m \sim E_{\text{Na}}$

\hookrightarrow approximated by top of AP

- when conductances equal, becomes "average" equation:

$$g_{\text{Na}} = g_{\text{K}} \quad E_{\text{Na}} + E_{\text{K}} / 2$$

- comes directly from equivalent circuit model

what is difference from E_{rev} ?

- E_{rev} when channel conducts 2 ions will also use this equation (eg AChR)

#4b from 2003:

↓ voltage clamp

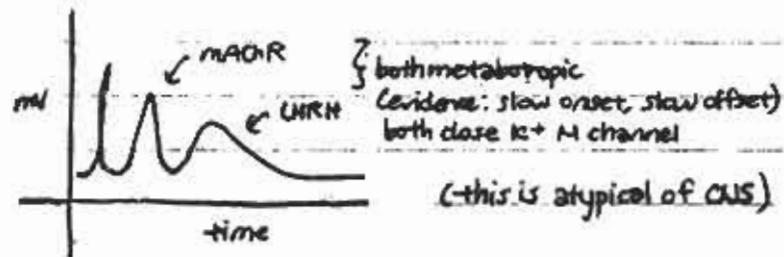
- put electrode in cell, inject current to pull away from resting, fire synapse, see if V goes up or down (which way current injected)
- or, patch-clamp, put on GABA

- know list of drugs for nAChR + mAChR; α -adrenergic, β -adrenergic

don't need to know particular names for these drugs

- if not covered in class, don't need to know

occlusion experiments



- eg in sympathetic ganglion, w/ fast, slow, late slow EPSPs

- do slow & late slow signaling events represent convergent pathways?

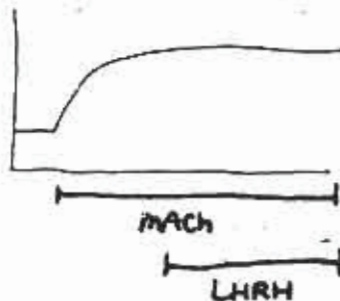
- yes, b/c affect same K^+M channel

- how do we find out? iontophorese for drug substance, record postsynaptically

- eg iontophorese muscarine; at some point during depolarization, iontophorese LHRH

- if no response, response to muscarine "occludes" response to LHRH

- then do in other order, to show that there is LHRH response (just no additional response)



equation for discharging capacitor: not for AP

- know equations for charging + discharging (need to know to solve # on Problem Set #1)

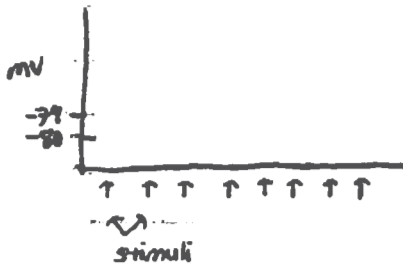


* will probably get questions w/ differentiated definition of capacitance: $\frac{dV}{dt} = -\frac{I}{C}$

- flow of positive charge inward = negative current, but depolarizing, so gives positive ΔV (that's why negative sign there)

- find slope, figure out current, area of capacitor (this type of question)

quantal analysis - bathe in low $[Ca^{2+}]$ solution to get inefficient transmission



- Stimulate over & over, get eg 1 mV deflections

2 pieces of information:

1. transmission quantized

2. statistically wobbly (1, 2, 3)

- if you get eg 5 out of 10 failures (lack of postsynaptic response), $P_0 = 50\%$

- Poisson distribution based on assumption of lots & lots of vesicles (like NHT); however, CNS synapses have much fewer synapses (in this case, use binomial distribution instead)

- eg 4 vesicles at CNS synapse each w/ 50% probability of release, $P_0 = \frac{1}{8}$

(Poisson only true if many vesicles)

always inversely related

- if increase probability of vesicle being released, increase m , decrease P_0

shows presynaptic event

- quantal analysis: look for change in P_0 for presynaptic effect

channels:

[α -helices (no polines) (hydrophobic A.A.s (eg valine, isoleucine))

1. voltage-gated: 6 TM, 4th (S4) not α -helix: every 3rd A.A positively charged

2. ligand-gated

↳ this is what moves in response to voltage change

3. 2nd messenger

↳ by crystallography, S4 turns out not to be α -helix; rather, is paddle out in membrane that flips outward w/ depolarization

(4 S4s per channel)

H & H wet problems:

- spatial changes in voltage (solve w/ space clamp so every patch of membrane at same V): gives you membrane AP (each patch goes off at same time: velocity of propagation infinite)
- need voltage clamp to avoid changes in conductance
- need way to separate I_{Na} & I_K (drugs) (or bath-changing experiments)

↳ used all the time by physiologists

changes gradients / resting potential, so eg can see effect of ion on resting potential, overshoot, E_{rev} , etc

- at Nernst equilibrium, energy lost by going down concentration gradient compensated by energy gained by going up voltage gradient

- effect of changing $[Cl^-]$?

- Cl^- not pumped (in class examples anyway) so does not have effect on resting V_m

$$V_{Cl} = -58mV \cdot \log \frac{[Cl^-]_o}{[Cl^-]_i}$$

- if adjusts itself passively, V_{Cl} will be resting V_m

So V_d is constrained; what you solve for is $\frac{[Cl^-]_o}{[Cl^-]_i}$

(look like silent synapses but are inhibitory)

- drugs that change both m & \bar{v}_i ? not in scope of course (but eg dull pores in membranes)

- dendrites poor in voltage-gated K^+ & Na^+ channels; no regenerative positive feedback response (this is why synaptic potentials graded while APs not)

permeability vs. conductance:

- don't need to know about permeability
- permeability is purely property of membrane; conductance is property of whole circuit

- net flow of ions at resting V_m ; so slight leakage of K^+ outward, tiny leakage of Na^+ inward; these are equal, so steady V

- would run down if not for $Na^+ K^+$ pump (this counteracts)

advantages of Aplysia: big cells, reproducible networks

- know habituation, sensitization

↳ mimicked by 5-HT application (upstream of PKA)

- know capacitive current in voltage clamp

- change changing of V a whole lot: if $\frac{dV}{dt}$ big, I is big

collagenase - lets you pull presynaptic & postsynaptic sides apart

- degrades collagen

Morris water maze - need hippocampus to perform well

(knock out both short-term & long-term memory)

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